

# Regulation of muscle size in humans: Role of myostatin?

**M.M. Bamman**

Associate Professor, Department of Physiology and Biophysics Director,  
Core Muscle Research Laboratory, University of Alabama at Birmingham School of Medicine

**Keywords:** Satellite Cell, Myoblast, Myostatin, IGF-I, Hypertrophy, Atrophy

Significant advances over the past few years have led to an improved understanding of the cellular and molecular processes regulating muscle mass and myofiber size. Much like bone, skeletal muscle is a highly plastic tissue exquisitely responsive to environmental cues including mechanical load, endocrine factors, neural activity, and autocrine/paracrine factors in the muscle microenvironment. As described by Dr. Esser, mechanical overload drives growth signaling in skeletal muscle. Conversely, a reduction in the nominal daily loading pattern or specific alterations in the muscle microenvironment can activate mechanisms leading to skeletal muscle atrophy as described by Dr. Kandarian. Many of the findings from animal model systems regarding muscle mass regulation have now been translated to adult humans. Through studies of muscle growth or regeneration, it has become increasingly apparent that key events driving developmental myogenesis recapitulate in adult myofibers in response to injury or increased mechanical load, and that successful myogenesis is dependent upon a complex array of co-ordinated activities regulating both net muscle protein synthesis and muscle stem (satellite) cell recruitment.

Autocrine/paracrine as well as endocrine factors (e.g., IGF-I) are important modulators of these processes. Myostatin, a member of the transforming growth factor  $\beta$  superfamily, is perhaps the single most powerful negative regulator of developmental myogenesis as demonstrated by marked muscle hypertrophy in homozygous mutant mice<sup>1</sup>, cattle<sup>2</sup>, and a single known human case<sup>3</sup>. This potent peptide "anti-growth" factor inhibits proliferation<sup>4,5</sup> and differentia-

tion<sup>6,7</sup> of myoblasts during development and evidence suggests it may also impair satellite cell function in differentiated adult muscle undergoing regeneration/growth<sup>8</sup>. Myostatin mRNA expression appears to be sensitive to mechanical load and other hypertrophy or atrophy stimuli. Its expression increases during multiple models of muscle atrophy<sup>9-14</sup> and, in fact, the magnitude of myostatin mRNA elevation is significantly related to the magnitude of type II myofiber atrophy consequent to disuse<sup>12</sup>. On the other hand, long-term resistance training<sup>15,16</sup>, as well as a single bout of resistance loading<sup>17,18</sup> markedly inhibit myostatin mRNA expression. While a physiologic consequence of altering myostatin mRNA in adult muscle has not been clearly demonstrated, the findings have sparked numerous efforts to suppress or block myostatin as a treatment for muscles suffering atrophy due to disuse, aging, or primary muscle myopathy<sup>19-22</sup>. It is abundantly clear that myostatin plays a dominant, negative role during mammalian muscle development but its role and importance in normal, healthy adult muscle remains poorly understood.

In this session, the speaker will overview some of the key metabolic and molecular processes regulating muscle size in adult humans with a focus on mechanisms driving skeletal muscle protein synthesis and muscle satellite cell recruitment. In the context of the latter, the state of the current literature on myostatin will be discussed including a recent application of K-means cluster analysis by the speaker's laboratory<sup>23</sup>.

## Learning objectives

The information presented in this seminar should enable you to better: (1) Understand the key metabolic and molecular processes regulating muscle size in humans; (2) Understand the biological actions of myostatin in skeletal muscle; (3) Appreciate the remarkable effects of myostatin inhibition in animal models; and (4) Compare and contrast the current evidence regarding the influence of myostatin in developing vs. adult muscle.

The author has no conflict of interest.

Corresponding author: *Marcas M. Bamman, Ph.D., Physiology and Biophysics, Core Muscle Research Laboratory, University of Alabama at Birmingham School of Medicine, MCLM 966, 1530 3rd Ave South, Birmingham, AL 35294-0005, USA*  
E-mail: *mbamman@uab.edu*

Accepted 11 August 2008

---

**References**

1. Benabdallah BF, Bouchentouf M, Tremblay JP. Improved success of myoblast transplantation in mdx mice by blocking the myostatin signal. *Transplantation* 2005;79:1696-1702.
2. Carlson CJ, Booth FW, Gordon SE. Skeletal muscle myostatin mRNA expression is fiber-type specific and increases during hindlimb unloading. *Am J Physiol* 1999;277:R601-6.
3. Dasarathy S, Dodig M, Muc SM, Kalhan SC, McCullough AJ. Skeletal muscle atrophy is associated with an increased expression of myostatin and impaired satellite cell function in the portacaval anastomosis rat. *Am J Physiol Gastrointest Liver Physiol* 2004;287:G1124-30.
4. Garma TM, Kobayashi CA, Haddad F, Adams GR, Bodell PW, Baldwin KM. Similar acute molecular responses to equivalent volumes of isometric, lengthening or shortening mode resistance exercise. *J Appl Physiol* 2007;102:135-43.
5. Haddad F, Adams GR. Aging-sensitive cellular and molecular mechanisms associated with skeletal muscle hypertrophy. *J Appl Physiol* 2006;100:1188-203.
6. Kim JS, Cross JM, Bamman MM. Impact of resistance loading on myostatin expression and cell cycle regulation in young and older men and women. *Am J Physiol Endocrinol Metab* 2005;288:E1110-9.
7. Kim JS, Petrella JK, Cross JM, Bamman MM. Load-mediated down-regulation of myostatin mRNA is not sufficient to promote myofiber hypertrophy in humans: a cluster analysis. *J Appl Physiol* 2007;103:1488-95.
8. Lalani R, Bhasin S, Byhower F, Tarnuzzer R, Grant M, Shen R, Asa S, Ezzat S, Gonzalez-Cadavid NF. Myostatin and insulin-like growth factor-i and -ii expression in the muscle of rats exposed to the microgravity environment of the Neurolab space shuttle flight. *J Endocrinol* 2000;167:417-28.
9. Langley B, Thomas M, Bishop A, Sharma M, Gilmour S, Kambadur R. Myostatin inhibits myoblast differentiation by down-regulating MyoD expression. *J Biol Chem* 277: 2002;49831-40.
10. McCroskery S, Thomas M, Maxwell L, Sharma M, Kambadur R. Myostatin negatively regulates satellite cell activation and self-renewal. *J Cell Biol* 2003;162:1135-47.
11. McPherron AC, Lawler AM, Lee SJ. Regulation of skeletal muscle mass in mice by a new TGF-beta superfamily member. *Nature* 1997;387:83-90.
12. McPherron AC, Lee SJ. Double muscling in cattle due to mutations in the myostatin gene. *Proc Natl Acad Sci USA* 1997;94:12457-61.
13. Reardon KA, Davis J, Kapsa RM, Choong P, Byrne E. Myostatin, insulin-like growth factor-1, and leukemia inhibitory factor mRNAs are upregulated in chronic human disuse muscle atrophy. *Muscle Nerve* 2001; 24:893-9.
14. Rios R, Carneiro I, Arce VM, Devesa J. Myostatin is an inhibitor of myogenic differentiation. *Am J Physiol Cell Physiol* 2002;282:C993-9.
15. Roth SM, Martel GF, Ferrell RE, Metter EJ, Hurley BF, Rogers MA. Myostatin gene expression is reduced in humans with heavy-resistance strength training: a brief communication. *Exp Biol Med* 2003;228:706-9.
16. Schuelke M, Wagner KR, Stolz LE, Hubner C, Riebel T, Komen W, Braun T, Tobin JF, Lee SJ. Myostatin mutation associated with gross muscle hypertrophy in a child. *N Engl J Med* 2004;350:2682-8.
17. Taylor WE, Bhasin S, Artaza J, Byhower F, Azam M, Willard DH Jr, Kull FC Jr, Gonzalez-Cadavid N. Myostatin inhibits cell proliferation and protein synthesis in C2C12 muscle cells. *Am J Physiol Endocrinol Metab* 2001;280:E221228.
18. Thomas M, Langley B, Berry C, Sharma M, Kirk S, Bass J, Kambadur R. Myostatin, a negative regulator of muscle growth, functions by inhibiting myoblast proliferation. *J Biol Chem* 2000;275:40235-43.
19. Tsuchida K. Activins, myostatin and related TGF-beta family members as novel therapeutic targets for endocrine, metabolic and immune disorders. *Curr Drug Targets Immune Endocr Metabol Disord* 2004;4:157-66.
20. Wehling M, Cai B, Tidball JG. Modulation of myostatin expression during modified muscle use. *FASEB J* 2000;14:103-10.
21. Whittemore LA, Song K, Li X, Aghajanian J, Davies M, Girgenrath S, Hill JJ, Jalenak M, Kelley P, Knight A, Maylor R, O'Hara D, Pearson A, Quazi A, Ryerson S, Tan XY, Tomkinson KN, Veldman GM, Widom A, Wright JF, Wudyka S, Zhao L, Wolfman NM. Inhibition of myostatin in adult mice increases skeletal muscle mass and strength. *Biochem Biophys Res Commun* 2003;300:965-71.
22. Zachwieja JJ, Smith SR, Sinha-Hikim I, Gonzalez-Cadavid N, Bhasin S. Plasma myostatin-immunoreactive protein is increased after prolonged bed rest with low-dose T3 administration. *J Gravit Physiol* 1999;6:11-5.
23. Zhu X, Hadhazy M, Wehling M, Tidball JG, McNally EM. Dominant negative myostatin produces hypertrophy without hyperplasia in muscle. *FEBS Lett* 2000;474:71-5.