

RANK, RANKL and OPG in inflammatory arthritis and periprosthetic osteolysis

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Abstract

Elucidation of the receptor activator of nuclear factor kappa B (RANK), its ligand (RANKL) and osteoprotegerin (OPG) as the final effectors of bone resorption has transformed our understanding of metabolic bone diseases and revealed novel therapeutic targets. Activation of the RANK-RANKL signaling pathway is directly responsible for dramatic focal erosions that are observed in inflammatory arthritis and aseptic loosening of orthopaedic implants. While these conditions share many features common to all metabolic bone disorders (e.g., osteoclastic resorption), they exhibit several unique properties, which are highlighted in this review. Most important is the relative inability of bisphosphonate therapy to inhibit osteolysis in joint inflammation and periprosthetic joint loosening and the unexpected effectiveness of anti-cytokine therapy in both rheumatoid and psoriatic arthritis. Herein, we provide a review of the role of RANK, RANKL and OPG in erosive arthritis and periprosthetic osteolysis and discuss the potential of anti-RANKL therapy for these conditions.

Keywords: Rheumatoid Arthritis (RA), Psoriatic Arthritis (PsA), Aseptic Loosening, Tumor Necrosis Factor-alpha (TNF α), Osteoclast Precursors (OCP)

Erosive arthritis

Rheumatoid arthritis (RA) is an autoimmune disease that affects approximately 1% of the population and is characterized by joint pain, stiffness and inflammation^{1,2}. Although the precise etiology of the disease remains elusive, various genetic, epigenetic and environmental factors are thought to play a role in the establishment and progression of RA.

It is now firmly established that the pro-inflammatory cytokine, tumor necrosis factor (TNF), is a central mediator of inflammation and matrix destruction in RA³. Although this review focuses on the role of RANK, RANKL and OPG in joint erosions, no discussion of RA would be complete

without highlighting the remarkable success of anti-TNF therapy in the treatment of rheumatoid and psoriatic arthritis. For their seminal role in defining TNF at the apex of the pro-inflammatory cytokine cascade and their pioneering work in anti-TNF clinical trials, Drs. M. Feldmann and R.N. Maini received the 2003 Lasker Clinical Medical Research Award⁴. Beyond these specific achievements, the success of anti-TNF therapy via injection of a recombinant soluble receptor (etanercept) or antibody (infliximab, adalimumab) personifies the ultimate goal of biotechnology and dramatically underscores how complex autoimmune diseases can be effectively suppressed using a targeted therapeutic approach.

A central feature of the RA is the highly osteodestructive process, which leads to three forms of bone loss: i) focal bone loss at the joint margins and in underlying subchondral bone (periarticular osteopenia); ii) localized resorption at the site of synovial attachment to bone (erosions); and iii) generalized osteoporosis involving the axial and appendicular skeleton⁵. Of particular interest is the local bone erosion because this radiographic manifestation reflects underlying disease activity, is a key outcome measure, and is associated with an unfavorable prognosis^{6,8}.

Anton Weichselbaum first described the focal bone erosions in RA joints as "caries of the joint ends" in 1878⁹. The histopathology of bone erosions is unique in that the lesions

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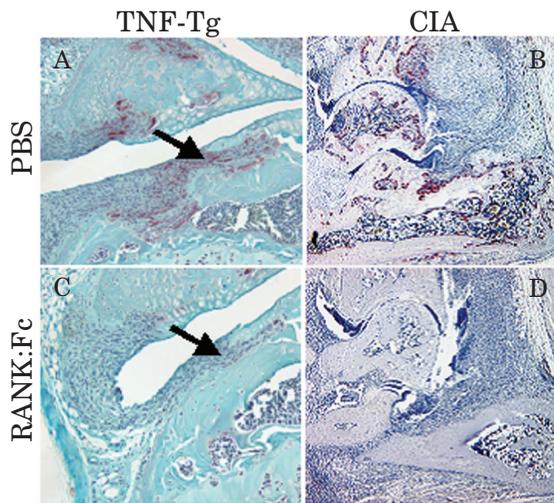


Figure 1. Efficacy of RANK blockade in mouse models of inflammatory arthritis. TNF-Tg mice (A+C) or mice with CIA (B+D) were treated with PBS (A+B) or high dose RANK:Fc (C+D). Histochemistry was performed on the knee joints (A+C) or ankles (B+D) by staining for TRAP (red). Arrows indicate the leading edge of the pannus, which mediates irreversible joint damage via subarticular erosions.

are eccentric and they emanate from the junction zone, where the bone, cartilage and synovial membrane are attached (Figure 1). Joint erosion is driven by the inflammatory synovial tissue or 'pannus', a hyperplastic, locally invasive tissue comprised of fibroblasts, monocytes and T lymphocytes, mast cells and numerous blood vessels. These cells produce vast array of inflammatory mediators, including cytokines (tumor necrosis factor (TNF)), interleukins (IL-1, IL-6, IL-17), prostaglandins (PGE₂), reactive oxygen species (O₂⁻, NO⁻), and matrix metalloproteinases (MMPs) that destroy the extracellular matrix in the joint by direct and indirect mechanisms. The pannus is extremely vascular, providing portals of entry for effector cells to enter the joint from the circulation and perpetuate joint destruction via autocrine and paracrine pathways.

Remarkably, it took more than a century for investigators to formally characterize the chondroclasts and osteoclasts positioned at the leading edge of the pannus tissue. These effector cells arise from different lineages and mediate focal bone erosions in the RA joint¹⁰. In seminal studies, Bromley and Woolley noted the very aggressive nature of these rapidly eroding lesions and they highlighted the presence of a bi-directional attack on the cartilage and bone¹¹, whereby the invading pannus drives "outside-in" erosions and cutting cones arising in the bone marrow erupt through the subchondral bone to cause "inside-out" erosions. Subsequent studies by Gravalles and colleagues further characterized the presence of osteoclast precursors (OCP) and mature osteoclasts within resorption lacunae of local bone erosions

by morphological features and molecular phenotype¹². From these central studies, the theory that osteoclasts are responsible for bone erosions in RA achieved broad acceptance.

RANK, RANKL and OPG in RA

Following the discovery that osteoclastogenesis and bone turnover are ultimately regulated by the expression of the osteoclast differentiation factor, RANKL vs its soluble inhibitor OPG^{13,14}, researchers began to investigate the expression of these molecules in tissues from RA patients. These early studies revealed that RANKL mRNA was detected by RT-PCR in whole synovial tissues from patients with RA but not in synovial tissues isolated from healthy controls. RANKL was also detected in cultured adherent synovial fibroblasts and activated T lymphocytes derived from RA synovial tissue¹⁵. In a parallel study, Takayanagi et al found that RANKL mRNA was highly expressed in all tissues from RA patients, but not from patients with osteoarthritis (OA)¹⁶. They also demonstrated that cultured rheumatoid synovial fibroblasts efficiently induce osteoclastogenesis in the presence of vitamin D, via up-regulation of RANKL and decreased OPG expression. In these studies, osteoclastogenesis was inhibited by OPG in a dose-dependent manner.

Several factors have been identified that increase the ratio of RANKL to OPG expression tipping the balance in favor of osteolysis but TNF has emerged as a dominant regulator. TNF directly stimulates RANKL production by stromal cells¹⁷, T lymphocytes¹⁸, B lymphocytes¹⁹, and endothelial cells²⁰. TNF also induces the expression of macrophage-colony stimulatory factor (M-CSF) by stromal cells²¹, which is the only other obligatory signal for osteoclastogenesis. TNF can promote RANKL expression by indirect mechanisms as well. For example, TNF-induced upregulation of prostaglandins, IL-1 or IL-17 can result in enhanced expression of RANKL²²⁻²⁴.

Psoriatic arthritis

Joint damage is also very common in psoriatic arthritis (PsA), an inflammatory joint disease that occurs in 10-15% of psoriasis patients²⁵. Gladman and colleagues noted that two-thirds of PsA patients manifest bone erosions radiographically on initial presentation to a rheumatologist²⁶. Of all the known arthropathies, PsA lesions are renowned for their marked destruction of cartilage and bone, particularly the arthritis mutilans subset reported in 16% of PsA patients²⁶. PsA joints often show extensive bone loss manifesting as eccentric erosions, frank tuft resorption and pencil-in cup deformities²⁷. Histopathologically, many PsA patients have aggressive synovitis with marked synovial hyperplasia, extensive vascular proliferation with a tortuous morphology and pannus tissue penetrating deep into cartilage and bone²⁸. In addition, osteoclasts are prominently situated at the bone-pannus junction and in bone marrow-

Model	Effect on Inflammation	Effect on Bone Erosion	Effect on Cartilage
Adjuvant arthritis in rats, OPG (1 mg/kg/day) ³¹	—	+++	+++
K/BxN (serum transfer) in RANKL ^{-/-} mice ³²	—	+++	+
CIA in rats, OPG (3 mg/kg/day) ³⁴	—	++	+
CIA in mice, RANK:Fc (10 mg/kg/48hr) ³⁶	—	+++	+
hTNF-Tg mice, OPG (6.4 mg/kg/day) ³⁷	—	++	—
hTNF-Tg mice, RANK:Fc (10 mg/kg/48hr) ⁴⁰	—	+++	+
hTNF-Tg x cFos ^{-/-} mice ³⁸	—	+++	—
hTNF-Tg x RANK ^{-/-} mice ⁴⁰	++	+++	+++

Table 1. Role of RANK signaling in animal models of inflammatory arthritis.

derived cutting cones traversing the bone matrix²⁹.

Immunohistochemistry revealed the very striking spatial regulation of RANK, RANKL and OPG expression in PsA joints²⁹. OPG expression was restricted to endothelial cells within the sub-synovial lining of the pannus. In contrast, intense RANKL immunoreactivity was identified in the outer lining of the synovium, where RANK positive monocytes, presumably osteoclast precursors (OCP) are also present. At the erosion front of the pannus, RANK positive multinucleated osteoclasts are found in resorption lacunae. Our interpretation of these data is that "outside-in" erosions occur as a result of OCP recruitment into the joint via the blood vessels in the synovium, where osteoclastogenesis is strongly inhibited by OPG. In response to the chemotactic cytokine stromal cell-derived factor-1 (SDF-1)³⁰, the OCPs migrate towards the RANKL-rich environment of the synovial lining, where they differentiate into active osteoclasts at the erosion front. In contrast, "inside-out" erosions are essentially void of all cell types except RANK positive monocytes and osteoclasts, and stromal cells. Presumably, these cells arise from precursors in the subchondral bone.

Lessons from animal models

Over the last decade, no technology has been more valuable to study the molecular pathogenesis of mammalian disease than genetically manipulated mice. By combining transgenic animals with highly selective protein inhibitors (antibodies and soluble receptors), rigorous *in vivo* gain and loss of function studies can be performed to elucidate the role of a particular gene in a disease state. Using established gene cloning methodologies and transgenic mouse models, investigators faithfully fulfilled Koch's postulates providing firm evidence that the RANK-RANKL signaling pathway is critical for osteoclast formation and bone resorption¹⁴.

Currently there are several well-established rodent models of inflammatory-erosive arthritis; and the roles of

RANK, RANKL and OPG have been investigated in many of them. These include adjuvant arthritis³¹, serum transfer³², collagen-induced arthritis³³⁻³⁶ and the TNF-transgenic mouse³⁷⁻⁴⁰. Immunohistochemistry and *in situ* hybridization studies revealed similar findings to the human studies described above. More importantly, genetic ablation and *in vivo* blockade experiments unequivocally demonstrated that RANK signaling is required for the genesis and progression of erosive arthritis. The results of these studies are summarized in Table 1 and can be visualized in Figure 1. In all cases, disruption of RANK signaling significantly inhibited osteoclast formation and/or induced osteoclast apoptosis, and prevented or inhibited erosion of cartilage and bone. The importance of NF κ B and AP-1 signaling immediately downstream of this RANK signal has also been confirmed in knockout mice¹⁴, as both signals are required for osteoclast formation and bone resorption. Thus, these pathways also serve as attractive targets of therapeutic intervention and are under extensive investigation. In contrast, no published study has demonstrated a significant role for this pathway in synovial inflammation, indicating that these two events are now separable at the molecular level. The view that inflammation and matrix destruction can be approached independently may have important therapeutic implications because pushing the doses of anti-inflammatory agents to lessen joint destruction can be immunosuppressive resulting in an increased risk for bacterial and opportunistic infections⁴¹.

Osteoclast Precursor (OCP) Frequency as a marker of erosive arthritis

One of the critical questions faced by physicians treating patients with RA is how aggressive to be with therapy in patients who present early in the disease course. In our pre-clinical^{40,42} and clinical²⁹ studies of TNF and RANK signaling in erosive arthritis we made several interesting observations indicating that OCP frequency in the blood may be a mark-

er of erosive disease including: i) TNF induces the release of CD11b+ OCP from the bone marrow into the circulation resulting in a significant increase in OCP frequency, by a mechanism independent of RANK signaling. ii) The frequency of circulating OCP correlated with the presence of erosive disease and, iii) anti-TNF therapy swiftly and dramatically reduced OCP frequency. Future studies designed to elucidate the mechanism of OCP release from the bone marrow, to fully characterize the surface marker phenotype of these OCPs and to better understand how they home to sites of focal erosions are underway. Ultimately, OCPs may be an important biomarker used to predict which patients with early RA are at greatest risk for joint damage and would benefit from aggressive therapeutic interventions.

The current frontier: chondroprotection and bisphosphonates

A central controversy in the field is the great disparity in findings regarding the chondroprotective effects of RANK blockade observed in the animal studies outlined in Table 1. Some studies documented that the articular surface of inflamed joints with extensive pannus were protected from erosion and proteoglycan loss as a result of RANK blockade, while other studies failed to observe these effects. The discrepancy in these studies is confounded by our lack of knowledge of the role of RANK and RANKL signaling in cartilage. Although the expression of RANK and RANKL in articular cartilage has been documented^{35,43,44}, experiments designed to demonstrate a functional role for these molecules in chondrocytes have failed to produce significant findings.

Another major controversy centers on whether bisphosphonates inhibit bone resorption in erosive arthritis. Based on the consistent efficacy of bisphosphonates in osteoporosis and the ability of these drugs to induce osteoclast apoptosis, scientists and clinicians predicted that they would be effective in preventing bone erosions in inflammatory arthritis. Although only a few clinical trials exploring this hypothesis have been published, the data were largely negative⁴⁵⁻⁴⁷. In more recent studies, investigators have demonstrated that etidronate therapy did not prevent radiologic progression in patients with RA⁴⁸, while others reported that etidronate significantly decreased Larsen damage scores in RA⁴⁹. In both studies etidronate decreased serum markers of bone turnover, suggesting a favorable effect on osteoporosis. The unimpressive results observed in clinical trials are consistent with animal studies that showed limited effects on erosion and osteoclast apoptosis in TNF-induced arthritis³⁷. In our *in vitro* experiments, we found that TNF protects osteoclasts from alendronate-induced apoptosis *in vitro*³⁶. We also observed that alendronate (10 mg/kg/day i.v. for 3 days) effectively induces OC apoptosis in the growth plate of hTNF-Tg mice, but OC in direct contact with synoviocytes at sites of focal bone erosion were unaffected. One explanation for these findings is that inflammatory cells residing in the pannus tissue deliver anti-apoptotic signals to osteoclasts at

the bone-pannus junction but not to those cells located in the growth plate. Collectively, these studies suggest that standard regimens of first generation bisphosphonates were largely ineffective in preventing focal bone erosions in inflammatory arthritis. Future trials with more potent bisphosphonates at higher doses than traditionally prescribed in osteoporosis are warranted to determine if these agents can significantly retard osteolysis in the inflamed joint.

Aseptic loosening of total joint replacements

Unfortunately, a common outcome of irreversible joint destruction from arthritis, trauma, cancer or avascular necrosis is loss of function and debilitating pain which can be alleviated by total joint replacement (TJR). Current estimates indicate that there are approximately 1.5 million TJR surgeries performed each year⁵⁰. The vast majority of procedures are for patients with severe osteoarthritis (OA) of the hip and knee. While TJR is considered to be one of the most successful surgical procedures in all of medicine, long-term outcomes are often limited by a condition known as "aseptic loosening". This condition, which takes 5-10 years to develop, is caused by chronic osteoclastic bone resorption around the implant until fixation is lost⁵¹. The current paradigm to explain aseptic loosening involves an inflammatory response to the wear debris particles produced by prosthetic implants⁵²⁻⁵⁵. These wear debris particles are phagocytosed by macrophages adjacent to the implant resulting in cell activation and the release of a diverse array of cytokines. This localized inflammatory response leads to the formation of a periprosthetic membrane with features similar to the synovitis of RA and PsA. Of particular interest, is the presence of osteoclasts that resorb bone at the bone-implant interface resulting in periprosthetic loosening.

RANK, RANKL and OPG in periprosthetic membranes

Periprosthetic membranes retrieved from patients with loose implants contain fibroblasts, macrophages, and a small number of T lymphocytes⁵⁶. As many as 10⁹ particles per gram of tissue can be recovered from periprosthetic membranes that are recovered during revision surgery^{53,57}. The inflamed tissues produce a variety of factors including TNF α , IL-1, IL-6, prostaglandin, and peptides that stimulate osteoclasts to resorb bone through the induction of RANKL^{56,58-61}. Since macrophages are the chief phagocytic cell ingesting wear debris particles, much attention has been focused on their role in cytokine production and osteoclast activation⁶²⁻⁶⁴. Indeed, macrophages located in the periprosthetic membrane are also OCP and, *in vitro* they differentiate into osteoclasts in response to: i) M-CSF and stromal cell derived factors⁶⁵, ii) RANKL alone⁶⁶, and iii) TNF and IL-1 in the absence of RANKL⁶⁷. Immunohistochemistry and *in situ* hybridization studies of periprosthetic membranes indi-

cate that macrophages are a source of RANKL in these tissues^{68,69}. Stromal cells and fibroblasts are also known to express this factor⁷⁰. Formal proof demonstrating the cellular source of the functional RANKL involved in periprosthetic osteolysis remains to be demonstrated.

Wear debris-induced osteolysis *in vivo*

While *in vitro* and *in situ* studies provide important information on the biology of aseptic loosening, *in vivo* experiments are critical to determine the true function of a specific pathway in this process. To this end, rat^{71,72} and canine⁷³ animal models have been developed to study wear debris-induced osteolysis. However, due to the availability of genetically defined mouse strains and the wealth of molecular probes, murine models have been most widely adopted⁷⁴⁻⁷⁷. Experimental studies in transgenic and knockout mice support the concept that wear debris particles stimulate osteolysis via NF κ B activation, and TNF production^{74,78,79}. Moreover, *in vivo* TNF blockade significantly inhibits but does not completely eradicate wear debris-induced osteolysis^{80,81}. In contrast, disruption of RANK signaling via genetic ablation or high dose RANK:Fc treatment (10 mg/kg/48hr) completely eliminates osteoclasts and bone resorption in this model⁸². Similar effects were also achieved via OPG gene therapy⁸³⁻⁸⁵. Of particular importance was the finding that new bone formation on both resorbed and unresorbed surfaces was not suppressed by complete osteoclast depletion via RANK:Fc treatment, and that this newly synthesized bone had normal mineral content and matrix composition⁸².

Towards a therapeutic intervention for aseptic loosening

Currently no therapies are approved for aseptic loosening. At this time clinical trial data on the effects of bisphosphonates in this condition are unavailable but unpublished reports and anecdotal evidence suggest that these agents are ineffective. In our view, the greatest limitation to the development of an effective intervention is the lack of an accurate and reliable outcome measure. Development of such an outcome measure is particularly challenging given that aseptic loosening takes years to develop and because progression is non-linear, small sampling of bone metabolites in urine or blood is not useful. Additionally, periprosthetic osteolysis involves a complex 3-dimensional lesion, rendering 1-dimensional (DEXA) or 2-dimensional (X-rays) radiology inaccurate. Therefore, we have developed 3-dimensional computerized tomography (3D-CT) methods to quantify periprosthetic osteolysis. We validated this approach by demonstrating the known correlation between polyethylene wear and osteolysis⁸⁶. Based on the success of this technique and the efficacy of TNF blockade in an animal model, we performed a clinical pilot to evaluate the efficacy of etanercept in 20 patients with established periprosthetic osteolysis⁸⁷. While

this study was not powered to evaluate drug efficacy, it concluded that the technique could determine a significant effect of a drug that inhibited osteolysis by 50% in a placebo controlled trial with 83 patients in each arm. Thus, future trials are now being proposed to develop the first therapeutic intervention for aseptic loosening.

Conclusions

Focal bone resorption in erosive arthritis and aseptic loosening remains a unique form of osteolysis that poses significant therapeutic challenges not observed in metabolic bone disorders. In particular, extensive focal bone loss arising from the adjacent dense hyperplastic inflammatory tissue and the relative resistance of these conditions to bisphosphonates necessitates development of alternative treatment strategies. The elucidation of RANK, RANKL and OPG as the final effectors of osteoclastogenesis and bone resorption is a true breakthrough providing invaluable insights into the mechanisms that underlie pathologic osteolysis. Formal proof that RANK and RANKL are viable targets in inflammatory osteolysis comes from studies in animal models of arthritis and wear debris-induced osteolysis showing that osteoclastogenesis and bone resorption do not take place void of RANK signaling *in vivo* and are significantly inhibited by RANK or RANKL blockade. While additional studies are required to determine the toxicity of agents that block molecules in the RANK-RANKL signaling pathway, preliminary studies in animal models suggest that they are well-tolerated. Moreover, the absence of major adverse events in phase I clinical trials of osteoporosis with recombinant OPG and anti-RANKL provide additional support for the tremendous potential of this innovative treatment strategy.

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References

1. Feldmann M, Brennan FM, Maini RN. Rheumatoid arthritis. *Cell* 1996; 85:307-310.
2. Firestein GS. Evolving concepts of rheumatoid arthritis. *Nature* 2003; 423:356-361.
3. Feldmann M, Maini RN. Anti-TNF alpha therapy of rheumatoid arthritis: what have we learned? *Annu Rev Immunol* 2001; 19:163-196.
4. Feldmann M, Maini RN. Lasker Clinical Medical Research Award. TNF defined as a therapeutic target for rheumatoid arthritis and other autoimmune diseases. *Nat Med* 2003; 9:1245-1250.
5. Goldring SR, Gravalles EM. Pathogenesis of bone erosions in rheumatoid arthritis. *Curr Opin Rheumatol* 2000; 12:195-199.
6. Arnett FC, Edworthy SM, Bloch DA, McShane DJ,

- Fries JF, Cooper NS, Healey LA, Kaplan SR, Liang MH, Luthra HS. The American Rheumatism Association 1987 revised criteria for the classification of rheumatoid arthritis. *Arthritis Rheum* 1988; 31:315-24.
7. Scott DL, Pugner K, Kaarela K, Doyle DV, Woolf A, Holmes J, Hieke K. The links between joint damage and disability in rheumatoid arthritis. *Rheumatology (Oxford)* 2000; 39:122-132.
 8. Van der Heijde D, Dankert T, Nieman F, Rau R, Boers M. Reliability and sensitivity to change of a simplification of the Sharp/Van der Heijde radiological assessment in rheumatoid arthritis. *Rheumatology (Oxford)* 1999; 38:941-947.
 9. Weichselbaum A. Die feineren Veränderungen des Gelenkknorpels bei fungöser Synovitis und Karies der Gelenkenden. *Archiv Pathol Anat Physiol Klin Med* 1878; 73:461-475.
 10. Bromley M, Woolley DE. Chondroclasts and osteoclasts at subchondral sites of erosion in the rheumatoid joint. *Arthritis Rheum* 1984; 27:968-975.
 11. Bromley M, Bertfield H, Evanson JM, Woolley DE. Bidirectional erosion of cartilage in the rheumatoid knee joint. *Ann Rheum Dis* 1985; 44:676-681.
 12. Gravalles EM, Harada Y, Wang JT, Gorn AH, Thornhill TS, Goldring SR. Identification of cell types responsible for bone resorption in rheumatoid arthritis and juvenile rheumatoid arthritis. *Am J Pathol* 1998; 152:943-951.
 13. Teitelbaum SL. Bone resorption by osteoclasts. *Science* 2000; 289:1504-1508.
 14. Boyle WJ, Simonet WS, Lacey DL. Osteoclast differentiation and activation. *Nature* 2003; 423:337-342.
 15. Gravalles EM, Manning C, Tsay A, Naito A, Pan C, Amento E, Goldring SR. Synovial tissue in rheumatoid arthritis is a source of osteoclast differentiation factor. *Arthritis Rheum* 2000; 43:250-258.
 16. Takayanagi H, Iizuka H, Juji T, Nakagawa T, Yamamoto A, Miyazaki T, Koshihara Y, Oda H, Nakamura K, Tanaka S. Involvement of receptor activator of nuclear factor kappaB ligand/osteoclast differentiation factor in osteoclastogenesis from synovio-cytes in rheumatoid arthritis. *Arthritis Rheum* 2000; 43:259-269.
 17. Hofbauer LC, Lacey DL, Dunstan CR, Spelsberg TC, Riggs BL, Khosla S. Interleukin-1beta and tumor necrosis factor-alpha, but not interleukin-6, stimulate osteoprotegerin ligand gene expression in human osteoblastic cells. *Bone* 1999; 25:255-259.
 18. Cenci S, Weitzmann MN, Roggia C, Namba N, Novack D, Woodring J, Pacifici R. Estrogen deficiency induces bone loss by enhancing T-cell production of TNF-alpha. *J Clin Invest* 2000; 106:1229-1237.
 19. Kanematsu M, Sato T, Takai H, Watanabe K, Ikeda K, Yamada Y. Prostaglandin E2 induces expression of receptor activator of nuclear factor-kappa B ligand/osteoprotegerin ligand on pre-B cells: implications for accelerated osteoclastogenesis in estrogen deficiency. *J Bone Miner Res* 2000; 15:1321-1329.
 20. Collin-Osdoby P, Rothe L, Anderson F, Nelson M, Maloney W, Osdoby P. Receptor activator of NF-kappa B and osteoprotegerin expression by human microvascular endothelial cells, regulation by inflammatory cytokines, and role in human osteoclastogenesis. *J Biol Chem* 2001; 276:20659-20672.
 21. Srivastava S, Toraldo G, Weitzmann MN, Cenci S, Ross FP, Pacifici R. Estrogen decreases osteoclast formation by down-regulating receptor activator of NF-kappa B ligand (RANKL)-induced JNK activation. *J Biol Chem* 2001; 276:8836-8840.
 22. Lubberts E, Joosten LA, Oppers B, Van Den Bersselaar L, Coenen-de Roo CJ, Kolls JK, Schwarzenberger P, Van de Loo FA, Van den Berg WB. IL-1-independent role of IL-17 in synovial inflammation and joint destruction during collagen-induced arthritis. *J Immunol* 2001; 167:1004-1013.
 23. Kotake S, Udagawa N, Takahashi N, Matsuzaki K, Itoh K, Ishiyama S, Saito S, Inoue K, Kamatani N, Gillespie MT, Martin TJ, Suda T. IL-17 in synovial fluids from patients with rheumatoid arthritis is a potent stimulator of osteoclastogenesis. *J Clin Invest* 1999; 103:1345-1352.
 24. Lubberts E, Joosten LA, Chabaud M, Van Den Bersselaar L, Oppers B, Coenen-De Roo CJ, Richards DC, Miossec P, Van Den Berg WB. IL-4 gene therapy for collagen arthritis suppresses synovial IL-17 and osteoprotegerin ligand and prevents bone erosion. *J Clin Invest* 2000; 105:1697-1710.
 25. Mease PJ. Tumor necrosis factor (TNF) in psoriatic arthritis: pathophysiology and treatment with TNF inhibitors. *Ann Rheum Dis* 2002; 61:298-304.
 26. Gladman DD, Farewell VT, Nadeau C. Clinical indicators of progression in psoriatic arthritis: multivariate relative risk model. *J Rheumatol* 1995; 22:675-679.
 27. Resnick D, Niwayama G. Psoriatic arthritis. In: Resnick D (ed) *Bone and Joint Imaging*. WB Saunders, Philadelphia; 1989:320-329.
 28. Fearon U, Griosos K, Fraser A, Reece R, Emery P, Jones PF, Veale DJ. Angiopoietins, growth factors, and vascular morphology in early arthritis. *J Rheumatol* 2003; 30:260-268.
 29. Ritchlin CT, Haas-Smith SA, Li P, Hicks DG, Schwarz EM. Mechanisms of TNF-alpha- and RANKL-mediated osteoclastogenesis and bone resorption in psoriatic arthritis. *J Clin Invest* 2003; 111:821-831.
 30. Yu X, Huang Y, Collin-Osdoby P, Osdoby P. Stromal cell-derived factor-1 (SDF-1) recruits osteoclast precursors by inducing chemotaxis, matrix metalloproteinase-9 (MMP-9) activity, and collagen transmigration. *J Bone Miner Res* 2003; 18:1404-1418.
 31. Kong YY, Feige U, Sarosi I, Bolon B, Tafuri A, Morony S, Capparelli C, Li J, Elliott R, McCabe S, Wong T, Campagnuolo G, Moran E, Bogoch ER, Van G, Nguyen LT, Ohashi PS, Lacey DL, Fish E, Boyle WJ,

- Penninger JM. Activated T cells regulate bone loss and joint destruction in adjuvant arthritis through osteoprotegerin ligand. *Nature* 1999; 402:304-309.
32. Pettit AR, Ji H, von Stechow D, Mueller R, Choi Y, Goldring SR, Benoist C, Gravallesse EM. TRANCE/RANKL knockout mice are protected from bone erosion in the K/BxN serum transfer model of arthritis. *Arthritis Rheum* 2001; 44 (Suppl.):1638.
 33. Romas E, Bakharevski O, Hards DK, Kartsogiannis V, Quinn JM, Ryan PF, Martin TJ, Gillespie MT. Expression of osteoclast differentiation factor at sites of bone erosion in collagen-induced arthritis. *Arthritis Rheum* 2000; 43:821-826.
 34. Romas E, Sims NA, Hards DK, Lindsay M, Quinn JW, Ryan PF, Dunstan CR, Martin TJ, Gillespie MT. Osteoprotegerin reduces osteoclast numbers and prevents bone erosion in collagen-induced arthritis. *Am J Pathol* 2002; 161:1419-1427.
 35. Lubberts E, Oppers-Walgreen B, Pettit AR, Van Den Bersselaar L, Joosten LA, Goldring SR, Gravallesse EM, Van Den Berg WB. Increase in expression of receptor activator of nuclear factor kappaB at sites of bone erosion correlates with progression of inflammation in evolving collagen-induced arthritis. *Arthritis Rheum* 2002; 46:3055-3064.
 36. Schwarz EM. Unpublished data.
 37. Redlich K, Hayer S, Maier A, Dunstan CR, Tohidast-Akrad M, Lang S, Turk B, Pietschmann P, Woloszczuk W, Haralambous S, Kollias G, Striner G, Smoiolen JS, Schett G. Tumor necrosis factor alpha-mediated joint destruction is inhibited by targeting osteoclasts with osteoprotegerin. *Arthritis Rheum* 2002; 46:785-792.
 38. Redlich K, Hayer S, Ricci R, David JP, Tohidast-Akrad M, Kollias G, Steiner G, Smolen JS, Wagner EF, Schett G. Osteoclasts are essential for TNF-alpha-mediated joint destruction. *J Clin Invest* 2002; 110:1419-1427.
 39. Schett G, Redlich K, Hayer S, Zwerina J, Bolon B, Dunstan C, Gortz B, Shultz A, Bergmeister H, Kollias G, Steiner G, Smolen JS. Osteoprotegerin protects against generalized bone loss in tumor necrosis factor-transgenic mice. *Arthritis Rheum* 2003; 48:2042-2051.
 40. Li P, Schwarz EM, O'Keefe RJ, Boyce BF, Xing L. RANK signaling independent and dependent stages of osteoclastogenesis in TNF-induced erosive arthritis. *J Bone Miner Res* 2003; 19:207-213.
 41. Ellerin T, Rubin RH, Weinblatt ME. Infections and anti-tumor necrosis factor alpha therapy. *Arthritis Rheum* 2003; 48:3013-3022.
 42. Li P, Schwarz EM, Li M, Looney RJ, Ritchlin CT, O'Keefe RJ, Boyce BF, Xing L. Systemic TNF α mediates an increase in peripheral CD11bhi osteoclast precursors in TNF α transgenic mice. *Arthritis Rheum* 2003; 50:265-276.
 43. Lacey DL, Timms E, Tan HL, Kelley MJ, Dunstan CR, Burgess T, Elliott R, Colombero A, Elliott G, Scully S, Hsu H, Sullivan J, Hawkins N, Davy E, Capparelli C, Eli A, Qian YX, Kaufman S, Sarosi I, Shalhoub V, Senaldi G, Guo J, Delaney J, Boyle WJ. Osteoprotegerin ligand is a cytokine that regulates osteoclast differentiation and activation. *Cell* 1998; 93:165-176.
 44. Komuro H, Olee T, Kuhn K, Quach J, Brinson DC, Shikhman A, Valbracht J, Creighton-Achermann L, Lotz M. The osteoprotegerin/receptor activator of nuclear factor kappaB/receptor activator of nuclear factor kappaB ligand system in cartilage. *Arthritis Rheum* 2001; 44:2768-2776.
 45. Ralston SH, Hacking L, Willocks L, Bruce F, Pitkeathly DA. Clinical, biochemical, and radiographic effects of aminohydroxypropylidene bisphosphonate treatment in rheumatoid arthritis. *Ann Rheum Dis* 1989; 48:396-399.
 46. Maccagno A, Di Giorgio E, Roldan EJ, Caballer LE, Perez Lloret A. Double blind radiological assessment of continuous oral pamidronic acid in patients with rheumatoid arthritis. *Scand J Rheumatol* 1994; 23:211-214.
 47. Eggelmeijer F, Papapoulos SE, Van Paassen HC, Dijkmans BA, Valkema R, Westedt JL, Landman JO, Pauwels EK, Breedveld FC. Increased bone mass with pamidronate treatment in rheumatoid arthritis. Results of a three-year randomized, double-blind trial. *Arthritis Rheum* 1996; 39:396-402.
 48. Valleala H, Laasonen L, Koivula MK, Mandelin J, Friman C, Risteli J, Kontinen YT. Two-year randomized controlled trial of etidronate in rheumatoid arthritis: changes in serum aminoterminal telopeptides correlate with radiographic progression of disease. *J Rheumatol* 2003; 30:468-473.
 49. Hasegawa J, Nagashima M, Yamamoto M, Nishijima T, Katsumata S, Yoshino S. Bone resorption and inflammatory inhibition efficacy of intermittent cyclical etidronate therapy in rheumatoid arthritis. *J Rheumatol* 2003; 30:474-479.
 50. Schwarz EM, Looney RJ, O'Keefe RJ. Anti-TNF α therapy as a clinical intervention for periprosthetic osteolysis. *Arthritis Res* 2000; 2:165-168.
 51. Harris WH. The problem is osteolysis. *Clin Orthop* 1995(311):46-53.
 52. Goldring SR, Jasty M, Roelke MS, Rourke CM, Bringhurst FR, Harris WH. Formation of a synovial-like membrane at the bone-cement interface. Its role in bone resorption and implant loosening after total hip replacement. *Arthritis Rheum* 1986; 29:836-842.
 53. Margevicius KJ, Bauer TW, McMahon JT, Brown SA, Merrit K. Isolation and characterization of debris in membranes around total joint prostheses [see comments]. *J Bone Joint Surg Am* 1994; 76:1664-1675.
 54. Horowitz SM, Doty SB, Lane JM, Burstein AH. Studies of the mechanism by which the mechanical failure of polymethylmethacrylate leads to bone resorption [see comments]. *J Bone Joint Surg Am* 1993; 75:802-813.
 55. Jacobs JJ, Shanbjag A, Glant T, Black J, Galante JO. Wear debris in total joint replacements. *J Am Acad*

- Orthop Surg 1994; 2:212-220.
56. Goldring SR, Schiller AL, Roelke M, Rourke CM, O'Neil DA, Harris WH. The synovial-like membrane at the bone-cement interface in loose total hip replacements and its proposed role in bone lysis. *J Bone Joint Surg Am* 1983; 65:575-584.
 57. Hirakawa K, Bauer TW, Stulberg BN, Wilde AH, Secic M. Characterization and comparison of wear debris from failed total hip implants of different types. *J Bone Joint Surg Am* 1996; 78:1235-1243.
 58. Goodman SB, Chin RC, Chiou SS, Schurman DJ, Woolson ST, Masada MP. A clinical-pathologic-biochemical study of the membrane surrounding loosened and non-loosened total hip arthroplasties. *Clin Orthop* 1989; 182-187.
 59. Jiranek WA, Machado M, Jasty M, Jevsevar D, Wolfe HJ, Goldring SR, Goldberg MJ, Harris WH. Production of cytokines around loosened cemented acetabular components. Analysis with immunohistochemical techniques and *in situ* hybridization [see comments]. *J Bone Joint Surg Am* 1993; 75:863-879.
 60. Chiba J, Rubash HE, Kim KJ, Iwaki Y. The characterization of cytokines in the interface tissue obtained from failed cementless total hip arthroplasty with and without femoral osteolysis. *Clin Orthop Rel Res* 1994; (300):304-312.
 61. Shanbhag AS, Jacobs JJ, Black J, Galante JO, Glant TT. Cellular mediators secreted by interfacial membranes obtained at revision total hip arthroplasty. *J Arthroplasty* 1995; 10:498-506.
 62. Glant TT, Jacobs JJ. Response of three murine macrophage populations to particulate debris: bone resorption in organ cultures. *J Orthop Res* 1994; 12:720-731.
 63. Nakashima Y, Sun DH, Trindade MC, Maloney WJ, Goodman SB, Schurman DJ, Smith RL. Signaling pathways for tumor necrosis factor-alpha and interleukin-6 expression in human macrophages exposed to titanium-alloy particulate debris *in vitro* [see comments]. *J Bone Joint Surg Am* 1999; 81:603-615.
 64. Blaine TA, Rosier RN, Puzas JE, Looney RJ, Reynolds PR, Reynolds SD, O'Keefe RJ. Increased levels of tumor necrosis factor-alpha and interleukin-6 protein and messenger RNA in human peripheral blood monocytes due to titanium particles. *J Bone Joint Surg Am* 1996; 78:1181-1192.
 65. Sabokbar A, Fujikawa Y, Neale S, Murray DW, Athanasou NA. Human arthroplasty derived macrophages differentiate into osteoclastic bone resorbing cells. *Ann Rheum Dis* 1997; 56:414-420.
 66. Itonaga I, Sabokbar A, Murray DW, Athanasou NA. Effect of osteoprotegerin and osteoprotegerin ligand on osteoclast formation by arthroplasty membrane derived macrophages. *Ann Rheum Dis* 2000; 59:26-31.
 67. Sabokbar A, Kudo O, Athanasou NA. Two distinct cellular mechanisms of osteoclast formation and bone resorption in periprosthetic osteolysis. *J Orthop Res* 2003; 21:73-80.
 68. Crotti TN, Smith MD, Findlay DM, Zreiqat H, Ahern MJ, Weedon H, Hatzinikolous G, Capone M, Holding C, Haynes DR. Factors regulating osteoclast formation in human tissues adjacent to peri-implant bone loss: expression of receptor activator NFkappaB, RANK ligand and osteoprotegerin. *Biomaterials* 2004; 25:565-573.
 69. Gehrke T, Sers C, Morawietz L, Ffernahl G, Neidel J, Frommelt L, Krenn V. Receptor activator of nuclear factor kappaB ligand is expressed in resident and inflammatory cells in aseptic and septic prosthesis loosening. *Scand J Rheumatol* 2003; 32:287-294.
 70. Sakai H, Jingushi S, Shuto T, Urabe K, Ikenoue T, Okazaki K, Kukita T, Jukita A, Iwamoto Y. Fibroblasts from the inner granulation tissue of the pseudocapsule in hips at revision arthroplasty induce osteoclast differentiation, as do stromal cells. *Ann Rheum Dis* 2002; 61:103-109.
 71. Howie DW, Vernon-Roberts B, Oakeshott R, Manthey B. A rat model of resorption of bone at the cement-bone interface in the presence of polyethylene wear particles. *J Bone Joint Surg Am* 1988; 70:257-263.
 72. Pap G, Machner A, Rinnert T, Horler D, Gay RE, Schwarzberg H, Neeumann W, Michel BA, Gay S, Pap T. Development and characteristics of a synovial-like interface membrane around cemented tibial hemiarthroplasties in a novel rat model of aseptic prosthesis loosening. *Arthritis Rheum* 2001; 44:956-963.
 73. Shanbhag AS, Hasselman CT, Rubash HE. The John Charnley Award. Inhibition of wear debris mediated osteolysis in a canine total hip arthroplasty model. *Clin Orthop* 1997(344):33-43.
 74. Merkel KD, Erdmann JM, McHugh KP, Abu-Amer Y, Ross FP, Teitelbaum SL. Tumor necrosis factor-alpha mediates orthopedic implant osteolysis. *Am J Pathol* 1999; 154:203-210.
 75. Schwarz EM, Benz EB, Lu AL, Goater JJ, Mollano AV, Rosier RN, Puzas JE, O'Keefe RJ. A quantitative small animal surrogate to evaluate drug efficacy in preventing wear debris-induced osteolysis. *J Orthop Res* 2000; 18:849-855.
 76. Kaar SG, Ragab AA, Kaye SJ, Kilic BA, Jinno T, Goldberg VM, Bi Y, Stewart MC, Carter JR, Greenfield EM. Rapid repair of titanium particle-induced osteolysis is dramatically reduced in aged mice. *J Orthop Res* 2001; 19:171-178.
 77. Wooley PH, Morren R, Andary J, Sud S, Yang SY, Mayton L, Markel D, Sieving A, Nasser S. Inflammatory responses to orthopaedic biomaterials in the murine air pouch. *Biomaterials* 2002; 23:517-526.
 78. Schwarz EM, Lu AP, Goater JJ, Benz EB, Kollias G, Rosier RN, Puzas JE, O'Keefe RJ. Tumor necrosis factor-alpha/nuclear transcription factor-kappaB signaling in periprosthetic osteolysis. *J Orthop Res* 2000; 18:472-480.

79. Clohisy JC, Teitelbaum S, Chen S, Erdmann JM, Abu-Amer Y. Tumor necrosis factor-alpha mediates polymethylmethacrylate particle-induced NF-kappaB activation in osteoclast precursor cells. *J Orthop Res* 2002; 20:174-181.
80. Childs LM, Goater JJ, O'Keefe, Schwarz EM. Efficacy of Etanercept for Wear Debris-Induced Osteolysis. *J Bone Miner Res* 2001; 16:338-347.
81. Childs LM, Goater JJ, O'Keefe RJ, Schwarz EM. Effect of anti-tumor necrosis factor-alpha gene therapy on wear debris-induced osteolysis. *J Bone Joint Surg Am* 2001; 83-A:1789-1797.
82. Childs LM, Paschalis EP, Xing L, Dougall WC, Anderson D, Boskey AK, Puzas JE, Rosier RN, O'Keefe RJ, Boyce BF, Schwarz EM. *In vivo* RANK signaling blockade using the receptor activator of NF- kappaB:Fc effectively prevents and ameliorates wear debris-induced osteolysis via osteoclast depletion without inhibiting osteogenesis. *J Bone Miner Res* 2002; 17:192-199.
83. Goater JJ, O'Keefe RJ, Rosier RN, Puzas JE, Schwarz EM. Efficacy of *ex vivo* OPG gene therapy in preventing wear debris-induced osteolysis. *J Orthop Res* 2002; 20:169-173.
84. Ulrich-Vinther M, Carmody EE, Goater JJ, Soballe K, O'Keefe RJ, Schwarz EM. Recombinant adeno-associated virus-mediated osteoprotegerin gene therapy inhibits wear debris-induced osteolysis. *J Bone Joint Surg Am* 2002; 84-A:1405-1412.
85. Yang SY, Mayton L, Wu B, Goater JJ, Schwarz EM, Wooley PH. Adeno-associated virus-mediated osteoprotegerin gene transfer protects against particulate polyethylene-induced osteolysis in a murine model. *Arthritis Rheum* 2002; 46:2514-2523.
86. Looney RJ, Boyd A, Totterman S, Seo GS, Tamez-Pena J, Cambell D, Novotny L, Olcott C, Martell J, Hayes FA, O'Keefe RJ, Schwarz E. Volumetric computerized tomography as a measurement of periprosthetic acetabular osteolysis and its correlation with wear. *Arthritis Res* 2002; 4:59-63.
87. Schwarz EM, Campbell D, Totterman S, Boyd A, O'Keefe RJ, Looney RJ. Use of volumetric computerized tomography as a primary outcome measure to evaluate drug efficacy in the prevention of peri-prosthetic osteolysis: a 1-year clinical pilot of etanercept vs. placebo. *J Orthop Res* 2003; 21:1049-1055.